



# Draft Genome Sequence of *Streptomyces* sp. Strain R1, Isolated from Water Canal Sediments, Possessing Antimicrobial and Plant Growth Promoting Capabilities

Neelma Ashraf,<sup>a,b</sup> Munir Ahmad Anwar,<sup>a</sup> Kalsoom Akhtar,<sup>a</sup> Sumera Yasmin,<sup>a</sup>  Shazia Khaliq<sup>a</sup>

<sup>a</sup>Industrial Biotechnology Division, National Institute for Biotechnology and Genetic Engineering (NIBGE), Constituent College of Pakistan Institute of Engineering and Applied Sciences (PIEAS), Faisalabad, Pakistan

<sup>b</sup>Chemical Ecology/Biological Chemistry, Department of Biology & Zukunftskolleg, University of Konstanz, Konstanz, Germany

**ABSTRACT** We present the genome sequence of *Streptomyces* sp. strain R1, isolated from water canal sediments and possessing genes responsible for antimicrobial metabolites and plant growth promotion. The genome assembly contains 7,936,694 bp with 72.24% of guanine-cytosine content. This genome will provide basic knowledge of the genes and pathways involved in the above mechanisms.

*Streptomyces* are soil-inhabiting, free living, Gram-positive filamentous bacteria that also present as symbionts of plants, insects, and animals (1). Due to their production of a broad range of bioactive secondary metabolites, *Streptomyces* have gained attraction in agriculture as biocontrol agents (2–9).

*Streptomyces* sp. strain R1 was isolated from water canal sediments from Faisalabad, Pakistan. 1 g of sample was suspended to serial dilution and cultured on casein starch peptone yeast-extract malt-extract (CSPY-ME) medium for 7 days at 30°C (10). A hard, powdery colony was selected and purified by restreaking onto a fresh plate. For genomic DNA extraction, a 5-day-old culture was subjected to a previously developed method with certain modifications (11). Three stainless steel beads were placed in a culture-containing Eppendorf tube and incubated at 37°C for 25 min after being rinsed with 120  $\mu$ L of an extraction solution that contained lysozyme (0.1 mg/mL). Proteinase-K (20 mg/mL) and RNase-A (0.1 mg/mL) were added, followed by a 5 min incubation at 65°C, to denature and remove proteins and RNA. The extracted DNA was resuspended in EB buffer with the same volume of solid-phase reversible immobilization (SPRI) beads (Bulldog Bio Inc., USA) for purification. A Quantit dsDNA HS assay (Thermo Fisher Scientific) was performed in triplicate to quantify the purified DNA using an Eppendorf AF2200 plate reader (Eppendorf UK Ltd., UK). The Nextera XT Library Preparation Kit (Illumina, San Diego, USA) was employed to prepare the genomic DNA libraries, making several modifications to the manufacturer's protocol due to the high guanine-cytosine content. In the polymerase chain reaction (PCR), 2 ng of DNA were used instead of 1 ng, and the elongation period was lengthened from 30 s to 1 min. For the DNA quantification and library preparation, Microlab STAR liquid handler (Hamilton Bonaduz AG, Switzerland) was used, and the Illumina libraries were created and quantified using a KAPA Library Quantification Kit (Roche LightCycler 96 quantitative PCR). Sequencing with 250 bp paired-end reads was done using an Illumina HiSeq 2500. Default parameters were employed for all specified "software".

Quality control was checked on pair end reads generating 178 $\times$  coverage. Trimmomatic 0.30 software was used to trim the adapted reads at a sliding window quality cutoff of Q15 (10, 12). For the variant calling, reads from the samples were separately aligned to references using bwa-mem 7.12 (13) under the default settings. A pileup file was generated from all of the alignments using SAMtools 1.9 (<https://sourceforge.net/projects/samtools/files/samtools/1.9/>) mpileup, again under the default settings. Variant matching was performed using VarScan

**Editor** J. Cameron Thrash, University of Southern California

**Copyright** © 2022 Ashraf et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Neelma Ashraf, neelma.ashraf@uni-konstanz.de, or Shazia Khaliq, skhaliq1976@gmail.com.

The authors declare no conflict of interest.

**Received** 16 July 2022

**Accepted** 2 August 2022

**Published** 16 August 2022

(version 2) (<http://dkoboldt.github.io/varscan/>) mpileup2cns under two different settings (for the two different output files), one with a 10% threshold (min-coverage, 3; min-var-freq, 0.1; *P* value, 0.05) and the other with a 90% threshold (min-coverage, 10; min-var-freq, 0.9; *P* value, 0.05), and this was annotated using snpEff (<http://pcingola.github.io/SnpEff/>). The *de novo* assembly was made with the help of SPAdes version 3.7 (14), using the default parameters, and contig annotation was carried out with Prokka 1.11 (15). A completeness analysis was performed by using the Microbial Genomes Atlas (MiGA) webserver (<http://microbial-genomes.org/>). The metrics were assembled using the Prokaryotic Genome Annotation Pipeline (PGAP) version 4.12 by NCBI ([https://www.ncbi.nlm.nih.gov/genome/annotation\\_prok/](https://www.ncbi.nlm.nih.gov/genome/annotation_prok/)).

*Streptomyces* sp. strain R1 has a high quality genome with a completeness of 99.1%, a genome size of 7,936,694 bp, a mean coverage of 178 $\times$ , a number of 3,125,371 reads, and a guanine-cytosine content of 72.24%. The genome assembly produced 113 contigs with an  $N_{50}$  value of 237,932 bp, with a length of 1,198,146 bp for the largest contig and 6,866 coding sequences with 67 tRNAs and 3 ncRNAs.

**Data accessibility.** The whole-genome sequencing project JAJIBA000000000, BioProject number PRJNA777709, SRA number SRX13087810, and BioSample number SAMN22870856 have all been deposited in GenBank. Furthermore, the accession number OL744553 was assigned for the 16S rRNA sequence in ENA/DDBJ/GenBank.

## ACKNOWLEDGMENTS

The research has been supported and funded by Zukunftscolleg under the ZUKOnnect Fellowship Program at the University of Konstanz. We thank Dieter Spittler from the University of Konstanz for providing access to the lab facilities and MicrobesNG for offering genome sequencing services.

The contributing authors have no conflicts of interest to report.

## REFERENCES

- Seipke RF, Kaltenpoth M, Hutchings MI. 2012. *Streptomyces* as symbionts: an emerging and widespread theme? *FEMS Microbiol Rev* 36:862–876. <https://doi.org/10.1111/j.1574-6976.2011.00313.x>.
- Colombo EM, Kunova A, Pizzatti C, Saracchi M, Cortesi P, Pasquali M. 2019. Selection of an endophytic *Streptomyces* sp. strain DEF09 from wheat roots as a biocontrol agent against *Fusarium graminearum*. *Front Microbiol* 1:2356.
- Kunova A, Cortesi P, Saracchi M, Migdal G, Pasquali M. 2021. Draft genome sequences of two *Streptomyces albidoflavus* strains DEF1AK and DEF147AK with plant growth-promoting and biocontrol potential. *Ann Microbiol* 71: 1–8. <https://doi.org/10.1186/s13213-020-01616-2>.
- Liu D, Yan R, Fu Y, Wang X, Zhang J, Xiang W. 2019. Antifungal, plant growth-promoting, and genomic properties of an endophytic actinobacterium *Streptomyces* sp. NEAU-S7GS2. *Front Microbiol*: 2077.
- Xiao K, Kinkel LL, Samac DA. 2002. Biological control of Phytophthora root rots on alfalfa and soybean with *Streptomyces*. *Biol Control* 23:285–295. <https://doi.org/10.1006/bcon.2001.1015>.
- Conn V, Walker A, Franco C. 2008. Endophytic actinobacteria induce defense pathways in *Arabidopsis thaliana*. *Mol Plant Microbe Interact* 21:208–218. <https://doi.org/10.1094/MPMI-21-2-0208>.
- Kurth F, Mailänder S, Bönn M, Feldhahn L, Herrmann S, Große I, Buscot F, Schrey SD, Tarkka MT. 2014. *Streptomyces*-induced resistance against oak powdery mildew involves host plant responses in defense, photosynthesis, and secondary metabolism pathways. *Mol Plant Microbe Interact* 27: 891–900. <https://doi.org/10.1094/MPMI-10-13-0296-R>.
- Suárez-Moreno ZR, Vinchira-Villarraga DM, Vergara-Morales DI, Castellanos L, Ramos FA, Guarnaccia C, Degrassi G, Venturi V, Moreno-Sarmiento N. 2019. Plant-growth promotion and biocontrol properties of three *Streptomyces* spp. isolates to control bacterial rice pathogens. *Front Microbiol* 10:290. <https://doi.org/10.3389/fmicb.2019.00290>.
- Olanrewaju OS, Babalola OO. 2019. *Streptomyces*: implications and interactions in plant growth promotion. *Appl Microbiol Biotechnol* 103:1179–1188. <https://doi.org/10.1007/s00253-018-09577-y>.
- Ashraf N, Bechthold A, Anwar MA, Khaliq S. 2020. Draft genome sequence of *Streptomyces* sp. strain BR123, endowed with broad-spectrum antimicrobial potential. *Microbiol Resour Announc* 9:e00972-20. <https://doi.org/10.1128/MRA.00972-20>.
- Nikodinovic J, Barrow KD, Chuck JA. 2003. High yield preparation of genomic DNA from *Streptomyces*. *Biotechniques* 35:932–936. <https://doi.org/10.2144/03355bm05>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Li H, Durbin R. 2009. Fast and accurate short read alignment with Burrows–Wheeler transform. *Bioinformatics* 25:1754–1760. <https://doi.org/10.1093/bioinformatics/btp324>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <https://doi.org/10.1093/bioinformatics/btu153>.