

Degradation of hydroquinone, gentisate, and benzoate by a fermenting bacterium in pure or defined mixed culture

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Abstract. From a methanogenic fixed-bed reactor fed with hydroquinone as sole energy and carbon source, a rod-shaped bacterium was isolated in pure culture which could degrade hydroquinone and gentisate (2,5-dihydroxybenzoate). In syntrophic coculture with either *Desulfovibrio vulgaris* or *Methanospirillum hungatei*, also benzoate could be degraded. Other substrates such as sugars, fatty acids, alcohols, and cyclohexane derivatives were not degraded. Sulfate, sulfite, or nitrate were not used as external electron acceptor. The isolate was a Gram-negative, non-motile, non-sporeforming strict anaerobe; the guanine-plus-cytosine content of the DNA was 53.2 ± 1.0 mol%. In pure culture, hydroquinone was degraded to acetate and benzoate, probably via an intermediate carboxylation. In syntrophic mixed cultures, all three substrates were converted completely to acetate. Phenol was never detected as a fermentation product.

Key words: Anaerobic degradation – Benzoate – Hydroquinone – Gentisate – Aromatic compounds – Carboxylation – Reductive dehydroxylation – Interspecies hydrogen transfer

Degradation of aromatic compounds under anaerobic conditions has been reported repeatedly since 1932 (for review, see Evans 1977; Sleat and Robinson 1984; Young 1984). Most of these studies dealt with degradation of aromatic compounds in sludge samples or undefined enrichment cultures. Pure culture studies have mostly been done with phototrophic, sulfate reducing, and denitrifying bacteria (Bak and Widdel 1986; Szewzyk and Pfennig 1987; Tschech and Fuchs 1987; Widdel 1988). By contrast, only few data are available on degradation of aromatic compounds by fermenting bacteria. Pure cultures of fermenting bacteria degrading trihydroxybenzenoids and trihydroxybenzoate have recently been described (Tsai and Jones 1975; Schink and Pfennig 1982; Krumholz and Bryant 1986), and degra-

dation of resorcinol to acetate and butyrate was reported for defined mixed cultures (Tschech and Schink 1985). Two bacteria have been described degrading benzoate in syntrophic coculture with a hydrogen-consuming bacterium (Mountfort et al. 1984; Tschech and Schink 1986). No fermenting bacterium degrading phenols has been isolated in pure culture, but degradability of these compounds under methanogenic conditions has been observed in enrichment cultures and sludge samples (Szewzyk et al. 1985; Young and Rivera 1985).

In this paper, we report on the isolation of a hydroquinone- and gentisate-fermenting bacterium which also catabolized benzoate when grown in coculture with either a sulfate-reducing or a methanogenic bacterium.

Material and methods

Source of organisms

Strain HQGö1 was isolated from the enrichment culture HQGö described in an earlier study (Szewzyk et al. 1985), which was originally inoculated with anoxic digester sludge from the municipal sewage plant in Göttingen, FRG.

Media and growth conditions

The freshwater medium used for cultivation and isolation was essentially the same as described previously (Widdel and Pfennig 1981; Schink and Pfennig 1982). Trace element solution SL10 (Widdel et al. 1983), selenite/tungstate solution (Tschech and Pfennig 1984), 6 vitamin solution and vitamin B12 solution (Pfennig 1978) were added from sterile stock solutions to the medium after autoclaving. Hydroquinone, sodium benzoate and sodium gentisate were added from filter-sterilized stock solutions which were kept under N₂ atmosphere. In addition to sulfide, dithionite was added (about 100 µmol/l) as a further, strong reducing agent before inoculation. Strain HQGö1 was isolated using the deep agar culture technique (Pfennig 1978). Purity was checked by microscopic examination and by growth tests in complex medium (AC medium; Difco Laboratories, Detroit, MI, USA).

Growth tests were carried out in duplicates at 28°C in liquid cultures in 20 ml screw cap tubes. Growth was followed by measuring the turbidity in a Spectronic 70

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spectrophotometer (Bausch & Lomb, Rochester, NY, USA). Several substrates were tested for degradation in the presence of sulfate and *Desulfovibrio vulgaris* strain Marburg as hydrogen scavenger.

Chemical analyses

Aromatic compounds were quantified by recording absorption spectra in diluted medium samples with a Gilford 250 spectrophotometer. For quantitation of benzoate which absorbs in the same wavelength range as sulfide, zinc acetate solution was added to the sample (100 µl of a 1 M Zn-acetate solution to 1.4 ml sample) before measurement to precipitate the H₂S. In addition, phenol was identified and quantified by gas chromatography (Schink and Pfennig 1982), benzoate by high-pressure liquid chromatography on an ultrasphere reversed-phase column (Beckman Instruments, München, FRG).

Fermentation products were identified and measured by gas chromatography as described earlier (Schink and Pfennig 1982). Sulfide was determined photometrically by the methylene blue method (Cline 1969).

Hydrogen in the culture medium was measured by gas chromatography with a detector based on the HgO-to-Hg vapour conversion technique (Seiler et al. 1980).

All chemicals used were of reagent grade purity and obtained from Merck, Darmstadt, FRG, and Fluka, Neu-Ulm, FRG.

Results

Isolation and cultivation

The hydroquinone-degrading strain HQGö1 was isolated from an anaerobic fixed bed reactor inoculated with a liquid enrichment culture which had been cultivated for more than two years with hydroquinone as sole energy and carbon source (Szewzyk et al. 1985). In this highly enriched mixed culture, a stable community had developed which consisted mainly of the hydroquinone-degrading bacteria and two types of methanogenic bacteria, namely, *Methanospirillum* sp. and *Methanothrix* sp. The same bacterial population was observed in the continuously operated fixed bed reactor, where it formed a thick biofilm of the filling material (manuscript in preparation). Isolation of the hydroquinone degrader was only possible if the redox potential in the deep agar cultures was lowered by addition of about 100 µM dithionite. Addition of yeast extract, rumen fluid, or casamino acids never stimulated growth of the hydroquinone-degrading bacterium in the agar cultures.

The isolate obtained after two deep agar dilution series was further cultivated in sulfide-prereduced freshwater medium with hydroquinone as sole energy and carbon source. Growth of the pure culture was optimal at 2 mM hydroquinone concentration. At higher concentrations, the lag-phase increased considerably, and no growth occurred at concentrations higher than 10 mM. Dithionite (100 µM) was always added to the growth medium before inoculation and was necessary for reliable growth. Stock cultures had to be transferred at intervals of 4–6 weeks, and were stored at room temperature. At temperatures below 10°C, cells lysed within 1–2 weeks.

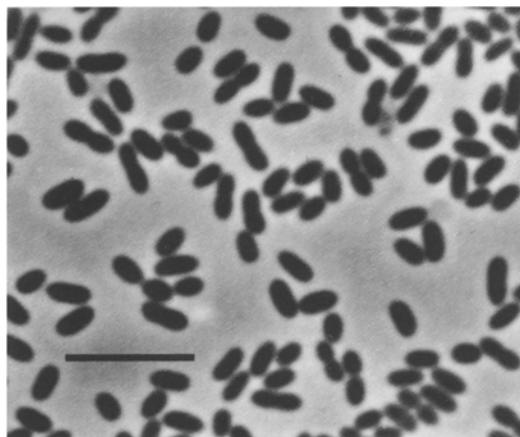


Fig. 1. Phase contrast photomicrograph of strain HQGö1 grown with hydroquinone. Bar equals 5 µm

Table 1. Substrates tested for growth of strain HQGö1. Substrate concentrations (in mmol/l) in the test cultures are given in parentheses. All growth tests were performed in duplicates

Substrates utilized:

Hydroquinone (2), benzoate (5)^a, gentisate (5).

Substrates not utilized:

- Formate (10), pyruvate (10), lactate (10), 3-hydroxybutyrate (10), malonate (10), succinate (10), malate (10), citrate (10), fumarate (10), caproate (10), acetoin (10), ethanol (10)
- Fructose (2), glucose (2), xylose (1), galactose (1)
- Phenol (0.5)^a, catechol (1), pyrogallol (1), phloroglucinol (1), resorcinol (1)
- Salicylate (1), gallate (2), 2,3-dihydroxybenzoate (1), nicotinate (1), protocatechuate (1)
- Cyclohexane carboxylate (1)^a, cyclohexanone (1)^a, cyclohexanol (1)^a
- Arbutine (2)

^a To these test tubes sulfate (10 mM) and *D. vulgaris* were added

Characterization of strain HQGö1

Cells of strain HQGö1 were short rods, 0.8 × 1.3–1.6 µm in size, with rounded ends, often in pairs (Fig. 1). Formation of longer cell filaments (10–15 µm) was rarely observed. Cells were always nonmotile, and no spores were formed. The Gram staining reaction was negative. The guanine-plus-cytosine content of the DNA was 53.2 ± 1.0 mol%. The optimal growth temperature was around 28°C; no growth was observed above 33°C and below 10°C.

Physiology

The only substrates degraded by strain HQGö1 in pure culture were hydroquinone and gentisate. In mixed cultures with either a sulfate-reducing or a methanogenic bacterium, also benzoate could be degraded (Table 1). Pure cultures converted hydroquinone to acetate and benzoate, and only traces of hydrogen (0.2–0.5 µmol/l) were formed (Table 2). Phenol was never found as intermediate or end product.

In coculture with a hydrogen-scavenging bacterium, acetate was the only organic degradation product of all three substrates, together with CH₄ or H₂S (Table 3).

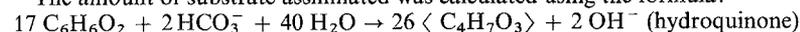
Table 2. Stoichiometry of substrate fermentation by strain HQGö1 in pure culture

Substrate	Amount of substrate provided (μmol)	Cell dry matter formed ^a (mg)	Substrate assimilated ^b (μmol)	Products formed		Cell yield ($\text{g} \cdot \text{mol}^{-1}$)	Electron recovery (%)
				Acetate (μmol)	Benzoate (μmol)		
Hydroquinone	200	3.0	19.0	390	58	14.9	103.3
Hydroquinone	300	4.4	27.9	590	82	14.7	101.5
Hydroquinone	500	7.1	45.0	960	145	14.3	101.7
Gentisate	300	5.6	35.5	570	78	18.5	100.3
Gentisate	500	9.1	57.7	945	135	18.2	101.0

All growth experiments were carried out at least in duplicates, in completely filled 100 ml screw cap bottles. All values given are means of at least two independent assays in every case.

^a The amount of cell matter formed was calculated from OD measurements via an experimentally determined conversion factor ($\text{OD}_{578} = 0.1 \cong 28.5 \text{ mg cell mass per l}$)

^b The amount of substrate assimilated was calculated using the formula:

**Table 3.** Stoichiometry of substrate fermentation by strain HQGö1 in mixed culture with *Desulfovibrio vulgaris* or *Methanospirillum hungatei*

Substrate	Amount of substrate provided (μmol)	Products formed (μmol)			Electron recovery ^a (%)
		Acetate	Sulfide	Methane	
Benzoate + <i>D. vulgaris</i>	500	1300	340	—	89.1
Benzoate + <i>M. hungatei</i>	250	700	—	175	93.5
Hydroquinone + <i>D. vulgaris</i>	250	630	50	—	83.7
Hydroquinone + <i>M. hungatei</i>	125	330	—	30	88.6

^a not including cell material

Experiments with *D. vulgaris* were carried out in completely filled screw cap bottles. Experiments with *M. hungatei* were carried out in serum bottles containing 50 ml medium and 55 ml headspace. All numbers are based on independent measurements of at least two replicate cultures in every case.

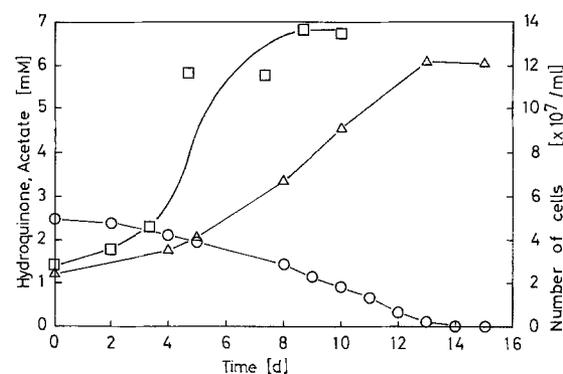


Fig. 2. Time course of hydroquinone fermentation (○) and formation of acetate (□) and cell material (●) by strain HQGö1. After the fifth day, the cells started to form clumps in formaldehyde-fixed slide preparations and exact cell counts were not possible any more.

The growth curve (Fig. 2) shows the dependence of growth and acetate formation on hydroquinone degradation in pure culture. 2.5 mM hydroquinone was completely degraded within 14 days, and 4.9 mM acetate, 0.7 mM benzoate, and 0.2–0.25 $\mu\text{mol H}_2/\text{l}$ were produced. The dependence of the hydroquinone degradation rate on the internal or external electron acceptor is shown in Fig. 3. The fastest

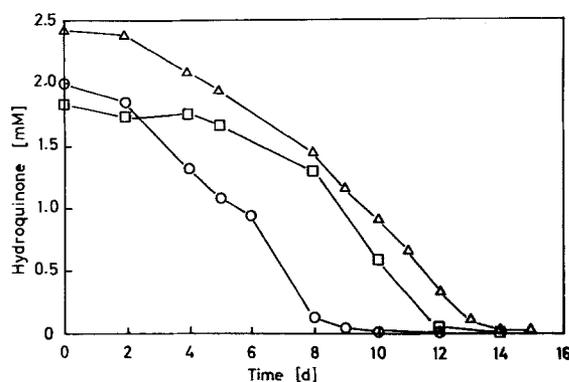


Fig. 3. Time course of hydroquinone degradation by strain HQGö1 with various electron acceptors. Δ pure culture, \square coculture with *Methanospirillum hungatei*, \circ coculture with *Desulfovibrio vulgaris*.

degradation was measured in coculture with *D. vulgaris* as partner (maximum degradation rate = 0.40 mM/day), whereas degradation in coculture with *M. hungatei* or in pure culture were lower (maximum degradation rate = 0.33 resp. 0.25 mM/day). Also benzoate degradation by cocultures was faster with *D. vulgaris* than with *M. hungatei* 0.50 mM benzoate/day versus 0.43 mM/day; data not shown).

as a fermentation product. The dominating non-methanogenic bacterium in the enrichment culture was morphologically similar to strain HQGö1. Therefore, phenol was probably formed in the enrichment culture by decarboxylation of salicylic acid by a further bacterium (Fig. 4). Our results obtained with the enrichment culture and the pure culture isolated thereof demonstrate again that conclusions on metabolic pathways in non-defined cultures should be drawn only very cautiously.

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