Generation and Characterization of Site-Specifically Mono-Ubiquitylated p53

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The tumor suppressor p53 is regulated by various posttranslational modifications including different types of ubiquitylation, which exert distinct effects on p53. While modification by ubiquitin chains targets p53 for degradation, attachment of single ubiquitin moieties (mono-ubiquitylation) affects the intracellular location of p53 and/or its interaction with chromatin. However, how this is achieved at the molecular level remains largely unknown. Similarly, since p53 can be ubiquitylated at different lysine residues, it remains unclear if the eventual effect depends on the position of the lysine modified. Here, we combined genetic code expansion with oxime ligation to generate p53 site-specifically mono-ubiquitylated at position 120. We found that mono-ubiquitylation at this position neither interferes with p53 ubiquitylation by the E3 ligases HDM2 and E6AP in complex with the viral E6 oncoprotein nor affects p53 binding to a cognate DNA sequence. Thus, ubiquitylation per se does not affect physiologically relevant properties of p53.

The growth-suppressive properties of the tumor suppressor p53 are negatively affected in the majority of human cancers by different means including mutations in the TP53 gene,[a] overexpression of the proto-oncoprotein HDM2, a member of the RING family of E3 ubiquitin ligases,[b] or mono-ubiquitylation at this position alters the property of p53 to serve as substrate for the E3 ligases HDM2 and E6AP.[c] To test whether this approach can also be applied to p53, we used the amber codon suppression (ACS) method to generate a p53 variant containing the pyrrolysine analog Plk at a distinct position and conjugated it via CuAAC to a substrate protein of interest.

We previously reported on the generation of defined Ub conjugates including Ub dimers and site-specifically mono-ubiquitylated forms of PCNA and the linker histone H1.2 by combining CuAAC with the genetic code expansion technology to incorporate unnatural amino acids at distinct positions of a protein of interest.[12a,c] To test whether this approach can also be applied to p53, we used the amber codon suppression (ACS) method to generate a p53 variant containing the pyrrolysine analog Plk at a distinct position and conjugated it via CuAAC to a Ub variant harboring azidohomoalanine at its C terminus. This resulted in reasonable amounts of p53-Ub conjugates (Figure S1 in the Supporting Information). We then subjected p53-Ub to ubiquitylation assays in the presence of HMD2 or the E3 ligase CHIP in complex with the viral E6 oncoprotein nor affects p53 binding to a cognate DNA sequence. Thus, ubiquitylation per se does not affect physiologically relevant properties of p53.
two E3 ligases. However, respective control reactions revealed that the reaction conditions of CuAAC, i.e. the presence of copper, drive p53 into a conformation that is neither recognized by HDM2 nor by the E6-E6AP complex (note that the structural requirements of p53 to serve as substrate for HDM2 and for the E6-E6AP complex are clearly different) (Figure S1).

p53 is a zinc-binding protein, and since copper does not interfere with ubiquitylation reactions per se (Figure S1C), we assumed that copper somehow affects the structural integrity of p53 as previously shown for other zinc-binding proteins. We therefore switched to oxime ligation, a bioorthogonal conjugation strategy that does not require metal ions and can be performed under non-denaturing conditions. In this reaction, an aminooxy group reacts with a carbonyl group to form an oxime linkage (Figure 1A). Furthermore, oxime ligation was used before to produce linkage-specific Ub conjugates as the oxime linkage is a reasonable mimic of the natural isopeptide bond (Figure S2). To equip p53 and Ub with the required functional groups at the desired positions, we used ACS again. In brief (for details, see Supporting Information), the lysine codon at position 120 of p53 was replaced by the amber stop codon enabling the introduction of a carbonyl group at this position via site-specific incorporation of ketosine (KeK). We chose this position, as K120 contributes to the sequence-specific DNA binding capacity of p53 and is known to be ubiquitylated, but whether K120 ubiquitylation affects DNA binding or other properties of p53 is unknown. E. coli cells harboring the respective expression plasmid and the required Pyrrolysine(Pyl)-tRNA-synthetase/tRNAPyl pair were cultured in presence of 10 mM KeK (resulting in the expression of p53-120KeK (Figure S3). Upon purification, the identity of p53-120KeK was verified by LC-MS/MS (Figure S6A).

To equip Ub with a C-terminal aminooxy group (Ub76ONH$_2$) by ACS, the C-terminal glycine codon at position 76 was replaced by the amber stop codon. As unnatural amino acid, we chose the lysine derivative N$_\varepsilon$-aminooxy-L-lysine, the aminooxy group of which had to be protected by an N$_\varepsilon$-latent protection group to prevent unwanted reactions with aldehyde or keto groups of endogenous bacterial molecules. As N$_\varepsilon$-latent protection group, we chose the acid-labile tert-butoxycarbonyl (Boc) group that also served as recognition motif for the respective Pyl-tRNA-synthetase/tRNAPyl pair for incorporation of N$_\varepsilon$-aminooxy-(Boc)-L-lysine (U1). Upon expression in E. coli and subsequent purification (Figure S4), Ub76ONH-Boc was deprotected by using 60% TFA, and the identity of Ub76ONH$_2$ was verified by LC-MS (Figure S4C). This also showed that incorporation of U1 did not proceed to completion (Figure S4C), resulting in a mixture of truncated Ub (Ub75) and full-length Ub (Ub76ONH-Boc) (Figure S4C). Although the truncated form cannot be separated from full-length Ub, this is not a concern since it cannot react with p53-120KeK. Thus, upon oxime ligation, Ub75 can easily be removed from the p53-Ub conjugate by anion exchange chromatography (for details, see Supporting Information).

To ensure that the functionality of p53 in the context of the envisioned p53-Ub conjugate is affected solely by the presence of Ub and not by other factors, the oxime ligation reaction should be performed under near-physiological conditions. However, oxime reactions proceed rather slowly at neutral pH. Since we previously showed that anionic surfactants enhance CuAAC bioconjugation reactions, we tested whether addition of non-denaturing concentrations of SDS also enhances oxime ligation. Indeed, increasing amounts of SDS resulted in increased yields of p53-120-Ub (Figure 1; Figure S4D, E) of up to 70% of the p53 input. That the ligation product indeed represents p53-120-Ub was verified by LC-MS/MS analysis (Figure S6B).

Upon further purification (Figure S5), we investigated the structural integrity of p53-120-Ub. Due to the fact that E6 recognizes p53 only when it is in a wild-type (wt)-like conformation, we determined whether the incorporation of KeK or the covalent attachment of Ub to position 120 affects

![Figure 1. Generation of mono-ubiquitylated p53 via oxime ligation. A) Schematic of the approach. For details, see text. B) Oxime ligation is enhanced with increasing amounts of non-denaturing SDS concentrations. p53-120KeK (20 pmol) was mixed with a 50-molar excess of Ub76ONH$_2$ in presence of increasing concentrations of SDS as indicated and subjected to two freeze-thaw cycles or, alternatively, to incubation for 20 h at 25 °C (Figure S4E). Reaction products were subjected to SDS-PAGE followed by Coomassie blue staining or by Western blot analysis using an anti-p53 antibody. Control, reaction in the absence of Ub76ONH$_2$. Running positions of p53-120KeK and p53-120-Ub are indicated by an arrow and an asterisk, respectively. Running positions of molecular mass markers (kDa) are indicated from top to bottom in B): 120, 100, 85, 70, 60, 50, 40, 30, 25, 20, 15, and in C): 70, 60, 50, 40.]
the ability of p53 to serve as a substrate in in vitro ubiquitylation assays using the E6-E6AP complex as E3 ubiquitin ligase (Figure 2A). Independent of the modification state, the p53 variants were recognized and ubiquitylated by the E6-E6AP complex to a similar extent. Similarly, like wt p53, p53-120KeK and p53-120-Ub were also ubiquitylated by HDM2 (Figure 2B). These data demonstrate that in contrast to CuAAC, the reaction conditions of oxime ligation do not affect the structural integrity of p53 and that mono-ubiquitylation at position 120 does not affect the property of p53 to be recognized by the E3 ligases tested.

Acetylation of p53 at K120 was reported to affect the sequence-specific DNA binding properties of p53.\(^{[21]}\) Therefore, we assessed the ability of p53-120KeK and p53-120-Ub to bind to the p21 response element (p21-RE) in electromobility shift assays (EMSA) in comparison to wt p53. Yet, no significant difference in the binding ability of the different p53 variants was observed (Figure 3A, Figure S7). While this further supports the notion that the reaction conditions of oxime ligation and the incorporation of KeK do not detectably affect the structural integrity of p53, it may seem surprising that attachment of a rather large molecule such as Ub to K120 does not interfere with DNA binding. However, it was recently reported that upon binding of p53 to so-called high affinity RE such as p21-RE, the side chain of K120 is disordered and not involved in hydrogen bonding with nucleobases.\(^{[24]}\) This provides a reasonable explanation for our observation that p53-120-Ub can still bind to the p21-RE and indicates that p53-120-Ub adopts a wt p53-like conformation.

Mono-ubiquitylation affects the intracellular localization of p53,\(^{[7–8]}\) which is likely due to changes in the protein-protein interaction properties of mono-ubiquitylated p53. Since cells or cell lysates are rich in de-ubiquitylating enzymes (DUBs), Ub conjugates that are resistant to the action of DUBs would be a prerequisite for such interactome analyses. To test the stability of oxime-linked p53-120-Ub, we incubated p53-120-Ub with rabbit reticulocyte lysate (RRL) for 30 min at 37°C. As previously shown for oxime-linked Ub-Ub conjugates\(^{[130]}\) and in contrast to enzymatically ubiquitylated p53, which served as control, the oxime-linked p53-120-Ub was not hydrolyzed. 150 ng oxime-linked p53-120-Ub or enzymatically mono-ubiquitylated p53 were incubated in the absence or presence of 5 μL rabbit reticulocyte lysate (RRL) for 30 min at 37°C. RRL served as a source of de-ubiquitylating enzymes. Reaction products were analyzed by SDS-PAGE followed by Western blot analysis with a p53-specific antibody. The running position of p53-120-Ub/ enzymatically mono-ubiquitylated p53 and the non-modified form of p53 is denoted with an asterisk and an arrow, respectively.

In conclusion, the combination of the genetic code expansion technology with oxime ligation enables the generation of mono-ubiquitylated forms of p53 and other proteins in a site-specific manner. Importantly, the reaction conditions of oxime ligation do not interfere per se with the structural integrity of p53 allowing to analyze known properties of p53 that may be affected by mono-ubiquitylation. Furthermore, oxime-linked p53-Ub conjugates are resistant to the action of

![Figure 3A](https://example.com/figure3a.png)

**Figure 3.** A) Characterization of the DNA binding ability of p53 variants by EMSA. Increasing concentrations of p53-120-Ub, p53-120KeK, and wild-type (wt) p53 were incubated with 10 nM fluorescein-labeled p21 response element (RE) and analyzed by fluorescence read out at 473 nm. - control reaction in the absence of p53; RE, running position of the unbound RE; ***, running position of the RE-p53 complex. B) Loading control for the proteins used in A). Running positions of non-modified p53/p53-120KeK and p53-120-Ub are indicated by an arrow and an asterisk, respectively. Running positions of molecular mass standards (kDa) are indicated on the left. C) The oxime-linkage of p53-120-Ub is not hydrolyzed. 150 ng oxime-linked p53-120-Ub or enzymatically mono-ubiquitylated p53 were incubated in the absence or presence of 5 μL rabbit reticulocyte lysate (RRL) for 30 min at 37°C. RRL served as a source of de-ubiquitylating enzymes. Reaction products were analyzed by SDS-PAGE followed by Western blot analysis with a p53-specific antibody. The running position of p53-120-Ub/enzymatically mono-ubiquitylated p53 and the non-modified form of p53 is denoted with an asterisk and an arrow, respectively.
DUBs. Thus, they are well suited for affinity enrichment experiments to identify proteins that interact selectively with monoubiquitylated forms of p53 thereby modulating the growth-suppressive properties of p53.

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Conflict of Interest

The authors declare no conflict of interest.

Data Availability Statement

The data that support the findings of this study are available in the supplementary material of this article.

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