

Does greater specific leaf area plasticity help plants to maintain a high performance when shaded?

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- **Background and Aims** It is frequently assumed that phenotypic plasticity can be very advantageous for plants, because it may increase environmental tolerance (fitness homeostasis). This should, however, only hold for plastic responses that are adaptive, i.e. increase fitness. Numerous studies have shown shade induced increases in specific leaf area (SLA), and there is wide consensus that this plastic response optimizes light capture and thus has to be adaptive. However, it has rarely been tested whether this is really the case.
- **Methods** In order to identify whether SLA plasticity does contribute to the maintenance of high biomass of plant species under shaded conditions, a meta analytical approach was employed. The data set included 280 species and 467 individual studies from 32 publications and two unpublished experiments.
- **Key Results** Plants increased their SLA by 55.4 % on average when shaded, while they decreased their biomass by 59.9 %. Species with a high SLA under high light control conditions showed a significantly greater ability to maintain biomass production under shade overall. However, in contrast to the expectation of a positive relationship between SLA plasticity and maintenance of plant biomass, the results indicated that species with greater SLA plasticity were less able to maintain biomass under shade.
- **Conclusions** Although a high SLA per se contributes to biomass homeostasis, there was no evidence that plasticity in SLA contributes to this. Therefore, it is argued that some of the plastic changes that are frequently thought to be adaptive might simply reflect passive responses to the environment, or result as by products of adaptive plastic responses in other traits.

Key words: Adaptive, functional traits, phenotypic plasticity, leaf mass area, LMA, low light environment, shade tolerance.

INTRODUCTION

Phenotypic plasticity is the ability of an organism to express different phenotypes in different environments, and is ubiquitous among organisms (Bradshaw, 1965, 1973; Schmid, 1992; West-Eberhard, 2003). Plants exhibit plasticity in numerous ecologically important traits related to plant function, development and life history (Sultan, 2000; Valladares *et al.*, 2007; Gratani, 2014). It is frequently assumed that phenotypic plasticity can be very advantageous for plants (Baker, 1974; Richards *et al.*, 2006), because it is thought to increase environmental tolerance (i.e. fitness homeostasis; Valladares *et al.*, 2014). This should, however, only hold for plastic responses that are adaptive, i.e. increase fitness (van Kleunen and Fischer, 2005; Muth and Pigliucci, 2007; van Kleunen *et al.*, 2011). Although many studies demonstrated that certain plastic responses of plants to contrasting environments are adaptive (Poorter and Lambers, 1986; Valladares and Pearcy, 1998; Donohue *et al.*, 2001), this is not always the case, as some plastic responses might also be neutral (i.e. do not affect fitness) or even maladaptive (i.e. decrease fitness; van Kleunen and Fischer, 2005; Sánchez-Gómez *et al.*, 2006; Ghalambor *et al.*, 2007). Therefore, it is important to assess explicitly whether the plasticity of a trait is adaptive

or not by investigating its contribution to the performance of plants in multiple environments.

Light, one of the crucial factors for the growth and development of plants, is a highly heterogeneous environmental resource in nature, and almost all plants are exposed to a certain degree of shading during their lifetime (Valladares and Niinemets, 2008). At low light intensity, photosynthesis and, consequently, plant growth, are reduced. Plants respond to changing light conditions by adjusting a suite of morphological and physiological traits, such as specific leaf area (SLA), internode and petiole lengths, leaf size, leaf thickness, leaf mass and chlorophyll content (Rozendaal *et al.*, 2006; Valladares and Niinemets, 2008; Legner *et al.*, 2014). While it is frequently implicitly assumed that these morphological and physiological changes are active plastic response to alleviate the effects on the plant of environmental stress, they could also reflect passive plastic responses to reduced resource availability (van Kleunen and Fischer, 2005).

Specific leaf area, the ratio of leaf area to leaf dry mass, is a key functional trait of plants underlying variation in growth rate among species (Pérez-Harguindeguy *et al.*, 2013). SLA is also a major trait in the worldwide leaf economics spectrum, which reflects the range of fast to slow returns on nutrient and dry

mass investment in leaves among species (Wright *et al.*, 2004; Flores *et al.*, 2014). Plants usually develop a higher SLA when grown under low-light conditions (Reich *et al.*, 2003; Rozendaal *et al.*, 2006; Feng and van Kleunen, 2014). This response could help plants to increase the efficiency of light capture and maximize carbon gain in such environments (Evans and Poorter, 2001; Gommers *et al.*, 2013), because SLA tends to scale positively with the mass-based light-saturated photosynthetic rate (Pérez-Harguindeguy *et al.*, 2013). Therefore, it is generally assumed that the plastic response of SLA enables plants to maintain a high performance under shading, and has to constitute adaptive plasticity (Valladares and Niinemets, 2008; van Kleunen *et al.*, 2011; Feng and van Kleunen, 2014). However, few studies have tested explicitly whether plastic responses to shading in SLA are really adaptive (but see Steinger *et al.*, 2003; Avramov *et al.*, 2006; Sánchez-Gómez *et al.*, 2006; McIntyre and Strauss, 2014 for notable exceptions), and thus result in a high performance of plants across different light intensities.

Here, we employed a meta-analytical approach to test whether plasticity of SLA in response to shading is adaptive, i.e. whether it enables plants to maintain their fitness under shade conditions. Fitness is ideally measured in terms of reproductive output; however, few studies have quantified this. Biomass is an alternative measure of plant performance, as it is the direct product of growth (e.g. Dawson *et al.*, 2012), and thus the change in biomass between high- and low-light conditions offers a good proxy for a species' ability to tolerate shade. We compiled a database of 467 studies from 32 publications and two unpublished experiments that measured the responses of biomass and SLA of 280 plant species to shading to test whether greater plastic changes in SLA in response to shading actually help the plants to better maintain performance under shade (i.e. whether plasticity in SLA is positively related to maintenance of plant biomass).

MATERIALS AND METHODS

Study and data collection

As a basis for the meta-analysis, we used a data set from a previous meta-analysis by Dawson *et al.* (2012), which was on the relationship between resource use and global naturalization success of plants. This data set included 15 studies on this topic published between 1990 and 2009. To obtain more recent studies (i.e. covering 2010–2014) on SLA and performance responses of plants to shading, we conducted a literature search in Web of Science (<http://apps.webofknowledge.com/>) using the following search string 'shad*' OR 'light*' OR 'R:FR' OR 'PAR' AND 'SLA' OR 'LMA' OR 'SLM'. In order to ensure that we did not miss any important studies, we also did a similar search in Google Scholar using the same keywords. Our searches resulted in 1055 new records. We then individually assessed each publication, and retained them if the study reported data on both plant biomass and SLA responses to shading. In total, we identified 33 publications that met our criteria (see [Supplementary Materials and Methods S1](#) for all publications used), covering 113 species and 280 individual studies. We also added unpublished data from two of our own experiments (D. Prati, unpubl. data; E. Haeuser, W. Dawson and M. van

Kleunen, unpubl. data) to the data set, yielding data on an additional 167 species and 187 individual studies.

We extracted mean values, sample sizes and measures of variance [i.e. s.d., s.e. or 95 % confidence intervals (CIs)] for plant biomass and SLA measures under a high-light control treatment and a shade treatment. We used the high-light treatment as the control treatment because we assumed it to be in the range of light intensities under which photosynthesis is light saturated. We did not consider studies that were performed in growth chambers with artificial lighting, because high-light conditions in growth chambers are much lower than in glasshouse and garden environments, and below the light intensity under which photosynthesis is light saturated. When more than one shading level was used for a single species, they were all included in our analyses (and compared with the same high-light control), but we accounted for multiple measurements per species in the analysis (see below). We extracted the data directly from the text or tables, or, when presented in figures, we extracted the data using the software Image J 1.47v (Rasband, 2013). We also extracted data on light intensity of the high-light control and shade treatments, and calculated the relative light intensity of the shade treatment compared with the control high-light treatment. Because light intensity in glasshouses is typically lower than that outdoors, we also extracted information on whether a study was conducted in a garden experiment or a glasshouse.

Effect size and variance

To examine the effects of shade treatment on SLA and plant biomass, we calculated the log response ratio ($\ln R$) as an effect size of response variables for each individual study following Hedges *et al.* (1999) as:

$$\ln R = \ln \left(\frac{X_s}{X_c} \right) = \ln(X_s) - \ln(X_c)$$

Here, X_s and X_c are the mean values of each individual SLA or biomass observation in the shade (S) and control (C) treatments, respectively. $\ln R$ values <0 indicate a decrease in SLA or biomass when shaded, and values >0 indicate an increase in SLA or biomass. The variance of $\ln R$ was, following Hedges *et al.* (1999), calculated as

$$v \ln R = \frac{(s.d._s)^2}{N_s(X_s)^2} + \frac{(s.d._c)^2}{N_c(X_c)^2}$$

Here, N_s , N_c , $s.d._s$, $s.d._c$, X_s and X_c are sample sizes, standard deviations and mean values for SLA or biomass in the shade (S) and control (C) treatments, respectively. As average biomass, and consequently also absolute changes in biomass in response to shading, might vary enormously among species (e.g. an annual herb has a much lower biomass than a tree), we chose the log response ratio as an effect size as it quantifies the proportional change instead of the absolute change in biomass (Hedges *et al.*, 1999).

Data analysis

All meta-analytical calculations and statistical analyses were performed in R 3.1.3 (R Core Team, 2015) using the package Metafor v1.9-5 (Viechtbauer, 2010). To test whether plastic changes in SLA in response to shading actually help the plant to better maintain performance (i.e. biomass) under shade, we selected a multivariate meta-analytical model using the `rma.mv` function. In the model, we included the effect sizes ($\ln R$) of biomass and their corresponding sampling variances as the response variable. As the main explanatory variable of interest, we included plasticity of SLA in response to shading (i.e. $SLA_{\text{shade}} - SLA_{\text{control}}$) in the model. Because the change in biomass may also depend on the SLA under high-light control conditions (SLA_{control}), we also included this baseline SLA as an explanatory variable in the model. Effectively, by including both SLA_{control} and $(SLA_{\text{shade}} - SLA_{\text{control}})$, we included both standard parameters (the intercept and slope) of a species linear SLA reaction norm to shading. We chose SLA under high-light conditions as the baseline (intercept) instead of SLA_{shade} , because the high-light conditions were likely to be more similar among studies than the low-light conditions. Moreover, while SLA_{shade} was strongly correlated with $(SLA_{\text{shade}} - SLA_{\text{control}})$ (Pearson $r = 0.812$, $P < 0.001$, $n = 467$), resulting in multicollinearity problems when including both variables in a single analysis, this was not the case for SLA_{control} and $(SLA_{\text{shade}} - SLA_{\text{control}})$ (Pearson $r = 0.084$, $P = 0.069$, $n = 467$), despite a strong correlation between SLA_{shade} and SLA_{control} (Pearson $r = 0.650$, $P < 0.001$, $n = 467$). As species varied in life form and studies varied in the degree of shading imposed, and in whether the study was done outdoors or in a glasshouse, we also included life form (woody vs. non-woody), relative light intensity (proportion of light in shade treatment compared with high-light control treatment) and experiment type (garden vs. glasshouse) as explanatory variables. The continuous explanatory variables ($SLA_{\text{shade}} - SLA_{\text{control}}$, SLA_{control} and relative light intensity) were all standardized by subtracting the mean and dividing by the s.d. for the entire data set, to facilitate interpretation and comparisons of the estimated model parameters (Schielzeth, 2010).

As effect sizes on the same species and from the same study are not independent, we included species and study as random factors. Moreover, as recent studies have shown that the addition of phylogenetic information could have a significant impact on the effect size estimates from meta-analysis models (Chamberlain *et al.*, 2012), we also included phylogenetic information as a variance covariance matrix in the model. We first constructed a base phylogenetic tree of all the species in our data set using the online program Phylomatic (Webb and Donoghue, 2005). Polytomies within this base tree were then solved as far as possible using published molecular phylogenies (see Supplementary Materials and Methods S2 for all publications used). The phylogenetic tree was transformed to an ultrametric tree using the `compute.brlen` function in the package `ape` v 3.2 (Paradis *et al.*, 2004). Finally, a variance covariance matrix was calculated from the ultrametric tree, representing phylogenetic relatedness among species, using the `vcv` function in the package `ape` v 3.2.

The estimates of effect size of biomass may be affected by whether or not the same genetic plant material is used in both

the high-light and shading treatments (Gianoli and Valladares, 2012) and by whether neutral shade (reduced light quantity alone) or canopy shade (reduced light quantity with altered spectral quality) is used (Griffith and Sultan, 2005). However, as in our data set, only six studies used the same genetic material in the different treatments and only three studies used canopy shade in high-light and shade treatments, we did not include these two factors in the main meta-analytical model described above. Instead, we performed separate analyses to test whether material used in each study (replicated genotype or non-replicated genotype) or shade type (neutral shade or canopy shade) had a significant influence on the estimates of the effect sizes of biomass and SLA in response to shading, using the `rma.mv` function. We included species and study in the model as random factors, and phylogeny as a variance covariance matrix. We also performed separate analyses to test whether experiment type (garden or greenhouse) or plant life form (woody or non-woody) had a significant influence on estimates of effect size of biomass and SLA in response to shading.

Using the models described above, we calculated a weighted mean effect size for each moderator. We calculated 95 % CIs with 1000 bootstrap replications, using the `boot.ci` function in the package `boot` v1.3-15 (Canty and Ripley, 2015). We considered the mean effect size estimate to be significantly different from zero if the 95 % CI around the mean did not include zero. In order to visualize the relationship between the plasticity of SLA and the changes in plant biomass in response to shading, we plotted all biomass effect sizes against SLA plasticity values, and added the regression line based on the predicted values from the main meta-analytical model described above. Total heterogeneity (Q_T) in the models used for separate analyses can be partitioned into heterogeneity explained by the model structure (Q_M) and unexplained heterogeneity (Q_E). We used the Q_M test to determine the significance of the difference in the mean effect size between different levels in the following moderator variables: plant material type (replicated genotype or non-replicated genotype), shade type (neutral shade or canopy shade), experiment type (garden or greenhouse) and plant life form (woody or non-woody). Because residual plots revealed a deviation from the assumption of normality, we used randomization tests to obtain a robust significance level of differences between groups (Q_M). By performing 1000 iterations for each model, a frequency distribution of possible Q_M values was generated. We then compared the randomly generated values with the observed Q_M value of each model, and calculated the proportion of randomly generated Q_M values more extreme (equal to or larger) than the observed Q_M values. We used this proportion as the significance level (i.e. P -value) for differences between groups.

RESULTS

On average, SLA of plants increased by 55.4 % when shaded, while biomass decreased by 59.9 % (Fig. 1). The responses of SLA and biomass to shading were not significantly affected by shade types (neutral or canopy), plant material type (replicated genotype or non-replicated genotype), experiment type (garden or greenhouse) or life form (woody or non-woody) (Fig. 1;

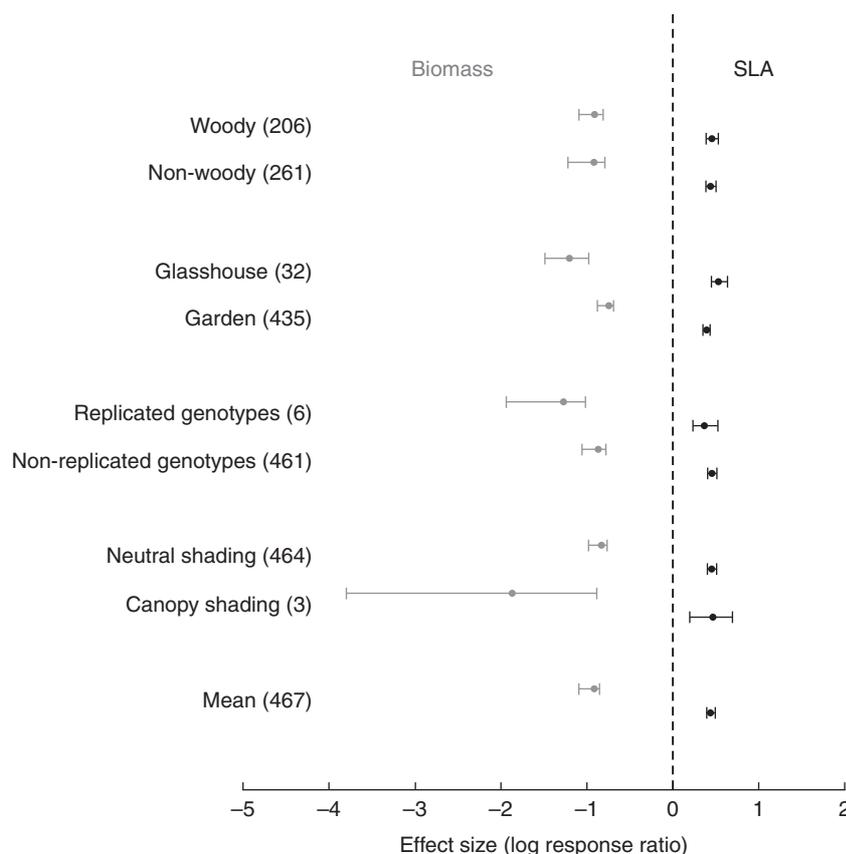


FIG. 1. Mean effect sizes (log response ratio) describing the overall responses of biomass and SLA to shading, and how these responses depend on whether the species are woody or non woody, and whether the study was done in a glasshouse or garden, used the same genetic material in the different light treatments, and used neutral or canopy shading. Error bars represent bias corrected bootstrapped 95 % confidence intervals around the mean effect size estimates derived from the phylogenetically corrected meta analytical model. The sample sizes (i.e. the number of studies) are given in parentheses. The dashed line indicates zero effect of shading.

Supplementary Data Table S1). The level of light in the shade treatment, relative to the high-light control treatment (mean, 41.5 %; range, 1–85.3 %) had no significant effect on the reduction in biomass (Fig. 2). Species with a greater SLA under control conditions (i.e. high light) showed a significantly smaller decrease in biomass under shade vs. control conditions overall (Figs 2 and 3). However, we found a negative relationship between $SLA_{\text{shade}} - SLA_{\text{control}}$ and $\ln R$ of biomass (Figs 2 and 3). In other words, the decrease in biomass under shading was significantly greater for plant species that showed a greater plastic increase in SLA. The variance component associated with phylogenetic history was low (0.0446), indicating that the effect sizes used in the analysis were not strongly phylogenetically related.

DISCUSSION

Specific leaf area is considered to be an important functional trait that may affect light interception and leaf longevity (Wright *et al.*, 2004), and is highly plastic in response to shading (Valladares and Niinemets, 2008). Although it is known that not all phenotypic plasticity increases performance (van Kleunen and Fischer, 2005), it is still frequently implied that plasticity in SLA should help plants maintain high performance

under varying light conditions (van Kleunen *et al.*, 2011; Gratani, 2014). Surprisingly, however, we found that greater plasticity of SLA of a species in response to shading was not associated with the maintenance of plant performance, but rather with greater reductions in plant biomass. Therefore, the results of our meta-analysis indicate that SLA plasticity to shading might not constitute adaptive plasticity.

Confirming the results of numerous previous studies on plant responses to shading (Reich *et al.*, 2003; Rozendaal *et al.*, 2006; Gianoli and Saldana, 2013; Feng and van Kleunen, 2014), our meta-analysis showed that most plants produced leaves with a higher SLA when shaded. This plastic response of SLA results in thinner, and relatively larger, leaves, and consequently should enhance light capture per gram of leaf tissue and thus mass-based photosynthesis. Therefore, it is frequently assumed that SLA plasticity represents adaptive shade tolerance plasticity, maximizing plant performance in the shade (Valladares and Niinemets, 2008; van Kleunen *et al.*, 2011; Freschet *et al.*, 2015). However, in contrast to support for this general assumption, we found a negative relationship between plant biomass responses to shading and SLA plasticity. In other words, our findings indicate that species that increased their SLA to a larger degree in response to shading were not more but less shade tolerant, compared with species that hardly changed their SLA.

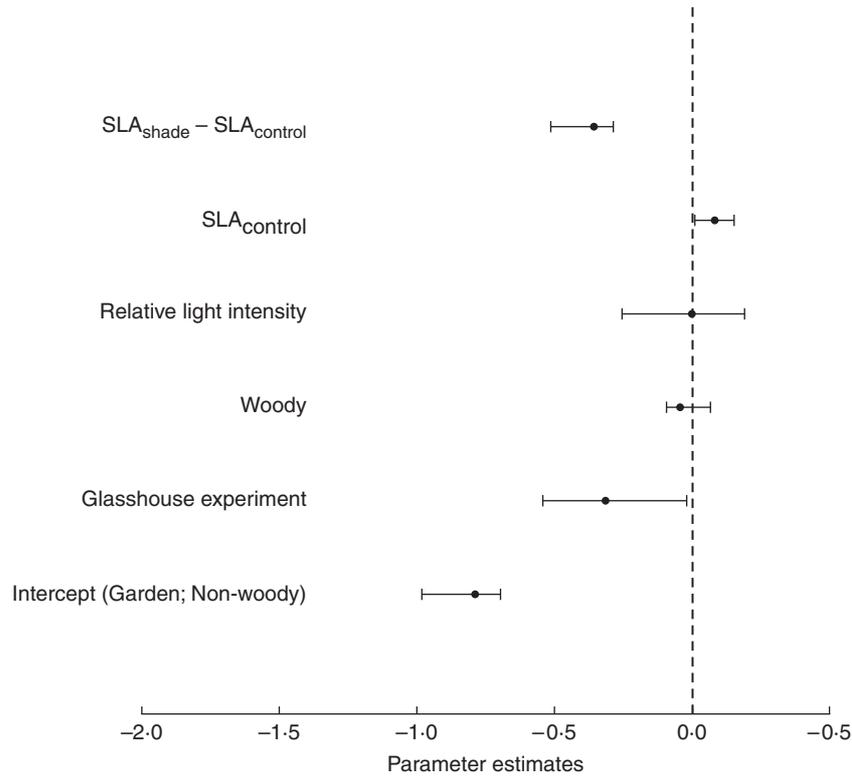


FIG. 2. Means of parameter estimates describing the relationship between biomass responses to shading [$\ln(\text{biomass}_{\text{shade}}/\text{biomass}_{\text{control}})$] and SLA plasticity in response to shading (i.e. $\text{SLA}_{\text{shade}} - \text{SLA}_{\text{control}}$), SLA in the high light control treatment ($\text{SLA}_{\text{control}}$), relative light intensity (percentage light in shade treatment relative to high light control treatment) and type of experiment (garden vs. glasshouse) on the changes of plant biomass in response to shading. Error bars show the bias corrected bootstrapped 95 % confidence intervals around the parameter estimates derived from the phylogenetically corrected meta analytical model. The dashed line indicates zero effect of the respective explanatory variable.

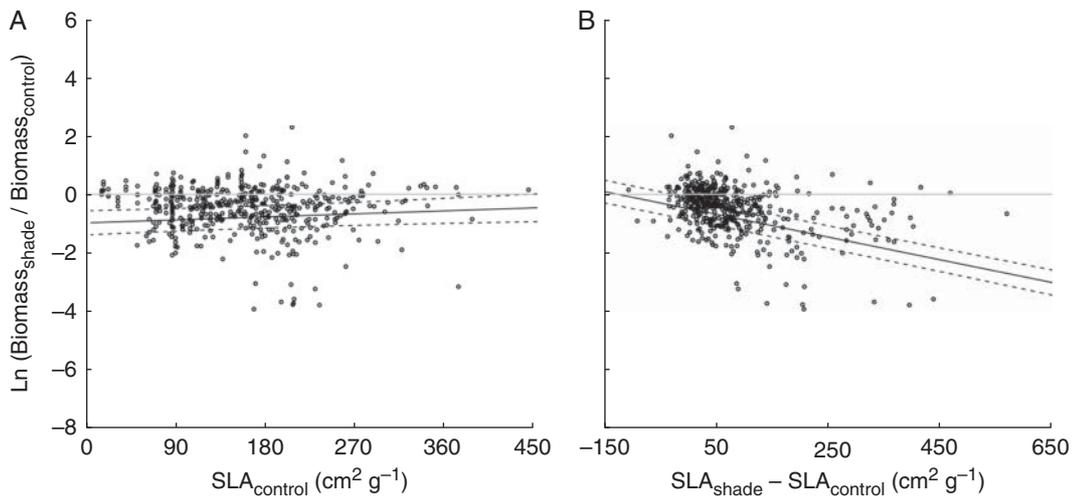


FIG. 3. Relationship between changes in plant biomass in response to shading, and (A) SLA in the high light control treatment ($\text{SLA}_{\text{control}}$, i.e. the intercept of the species' reaction norm) and (B) the changes in SLA (i.e. the slope of the species' reaction norm). The regression line is based on the predicted values from the phylogenetically corrected meta analytical model. The solid line is the fitted line, and the dashed lines are 95 % confidence intervals of the fitted line.

Few other studies have tested explicitly whether shade-induced responses in SLA are adaptive. Avramov *et al.* (2006) tested the adaptive value of plasticity in SLA of plants from two populations of *Iris pumila* grown at three light levels, and

found evidence that the plastic response in SLA to light availability was in the direction of values favoured by selection in one of the two populations (i.e. adaptive). Moreover, McIntyre and Strauss, (2014) investigated patterns of plasticity and

selection on SLA of *Claytonia perfoliata* plants grown in an oak canopy understorey and an adjacent grassland habitat, and found that *C. perfoliata* exhibited plastic responses in SLA in the same direction as promoted by selection (i.e. selection for a higher SLA in a canopy habitat), suggesting that the plastic response in SLA is adaptive. These two results thus contrast with the findings of our meta-analysis. One possible explanation for the discrepancy might be that these other studies tested for the benefit of plasticity within species, while we tested for the benefit of plasticity among species. Therefore, we clearly need more studies that assess the fitness effects of SLA plasticity in response to shading within species to see whether this plasticity is generally beneficial within species.

Our findings do not just suggest that a strong plastic increase in SLA of a species in response to shading is non-adaptive, but even suggest that it is maladaptive. One possible explanation could be that SLA plasticity is genetically and developmentally linked to plasticity in shade-avoidance traits, such as petiole and internode elongation. In contrast to a shade-tolerance trait, a shade-avoidance trait should help the plants to escape from the shade conditions by overtopping the neighbouring plants that impose the shade or by finding gaps in the vegetation. However, as most experiments on shade responses use artificial shading treatments from which the plants cannot escape, elongation responses are futile and might even be costly (Valladares *et al.*, 2007; Valladares and Niinemets, 2008). Another explanation for the negative association between SLA plasticity and biomass homeostasis could be that most studies measure SLA at the end of the experiment. If SLA determines light interception per gram of leaf, then plants that are able plastically to adjust SLA early should be able to maintain a high biomass production. However, SLA as measured at the end of an experiment might not be driving the performance of plants but might result from it. In other words, a plant that is not very shade tolerant, and thus shows a strong decrease in biomass in response to shading, will not have the resources (e.g. photo-assimilates) to produce thick leaves with a low SLA. A low SLA might be beneficial, also under shaded conditions, if it results in a greater proportion of incident photon capture per unit leaf area. Alternatively, it could be that plants do not actively increase their SLA in response to low light but instead passively decrease their SLA in response to high light due to accumulation of non-structural carbohydrates (thus increasing dry mass per leaf area) when the carbohydrate production exceeds the demand in meristems. Whatever the exact reason is for the negative association between SLA plasticity and biomass homeostasis, we recommend that future studies on this topic should measure SLA not only at the end of an experiment but also early on, and that they should impose more realistic shade treatments that allow shade-avoidance responses to be effective.

While our results indicate that SLA plasticity in response to shading is not adaptive, one could argue that our results indicate that SLA plasticity is adaptive in response to an increase in light intensity. In other words, if one uses the shade environment as the reference instead of the high-light environment, the plant species that have a stronger plastic decrease in SLA in response to high light are better able to take advantage of the high light intensity in terms of biomass production (Supplementary Data Fig. S1a). To gain more insight into the

underlying cause of the relationship between biomass change and SLA plasticity, we also carried out a regression of biomass in high- and low-light environments separately against SLA plasticity (Supplementary Data Fig. S1b). Plant biomass in high-light environments varied little in relation to SLA plasticity (Fig. S1b), but biomass under low-light environments decreased with increasing SLA plasticity (Fig. S1b). This indicates that species with greater SLA plasticity do not have an advantage under high-light conditions, but are disadvantaged under shade compared with less plastic species. In other words, the reduced ability of plants to produce biomass due to a lack of light in shaded environments is not compensated by increasing SLA to a greater degree, but is rather exacerbated by it.

Although SLA plasticity did not help plants to maintain a high performance when shaded, our results showed that species with greater SLA under high-light control conditions have a significantly smaller decrease in biomass when shaded. So, while plasticity in SLA did not increase biomass homeostasis, high SLA values did. Generally, shade-intolerant species have higher light compensation points and light-saturated photosynthetic rates (Givnish, 1988; Kitajima, 1994; Valladares and Niinemets, 2008), thus plants with high SLA values would be more shade tolerant. This finding supports the carbon-gain hypothesis, which states that any trait related to light-use efficiency that improves carbon gain in plants will increase performance under shade (Givnish, 1988; Valladares and Niinemets, 2008). Our finding is also in line with the many studies that found that species with a greater SLA are more shade tolerant (e.g. Sánchez-Gómez *et al.*, 2006; Janse-Ten Klooster *et al.*, 2007; Gianoli and Saldana, 2013). Although the relationship between the biomass response and SLA_{control} in our meta-analysis was shallow, it raises the question of why not all species have evolved greater SLA. Most probably, this is because some species do not encounter much shading in nature, and other selective forces, such as herbivory and drought stress, and environments favouring leaf longevity (Supplementary Data Fig. S2), have resulted in the evolution of species with low SLA. Additionally, while plants with lower SLA are less efficient in terms of metabolic cost per unit leaf area, they might capture a greater proportion of incident photons. When the increased photon capture more than offsets the increased metabolic cost of a lower SLA, the lower SLA should be favoured.

As species that naturally occur in shaded habitats are presumably more shade tolerant, it could be that the positive relationship between the change in biomass and SLA arose because species from shade habitats have higher SLA values than species from non-shade habitats. As information on the natural habitats is not available for most of the study species, we could not account for this in the main analysis. However, for 136 of the 280 study species, we had data on their Ellenberg light-indicator values (Ellenberg, 1974), which indicate the light conditions in the natural habitat of the species in Europe. Although this sub-set of species did not contain species from deep-shade habitats, we did not find evidence that species with different light-indicator values differed in SLA under high-light and under shaded conditions (Supplementary Data Fig. S3). Therefore, it is unlikely that our result of a higher biomass homeostasis for species with higher SLA values is confounded by species from shade habitats having higher SLA values.

Surprisingly, our results showed that relative light intensity had no significant effect on the reduction in biomass (Fig. 2). This runs counter to the results of many experiments, where biomass typically declines more or less continuously with declining light levels (e.g. Feng and van Kleunen, 2014; Kumar *et al.*, 2014; Konvalinková *et al.*, 2015). A likely explanation for this apparent discrepancy is that most species in our meta-analysis were not grown under more than two experimental light conditions, and that the light conditions varied among studies. Seventy of the 280 species were grown under more than two light levels, and a post-hoc analysis for this subset of species showed that within species, biomass declines more or less continuously with declining light levels (Supplementary Data Fig. S4). However, if we run the full meta-analytical model for this sub-set of 70 species, the effect of relative light intensity was still not significant, and the other results also remained qualitatively the same (Supplementary Data Fig. S5). So, while within each species relative light intensity is important for the change in biomass, among species it plays no significant role.

Conclusions

In summary, our meta-analysis suggests that plasticity in the ability of plants to capture more light per gram of leaf mass invested under low-light conditions by increasing SLA does not contribute to shade tolerance of plant species in terms of biomass homeostasis, and thus does not constitute adaptive phenotypic plasticity. This is despite wide consensus that plasticity in SLA and other traits associated with shade avoidance and tolerance, such as leaf length, leaf area, shoot root ratio, chlorophyll content and photosynthesis, can be adaptive (Dudley and Schmitt, 1996; Schmitt *et al.*, 1999; van Kleunen and Fischer, 2005; Valladares and Niinemets, 2008; van Kleunen *et al.*, 2011). We argue that some of the plastic responses of plant species to shade that are frequently thought to be adaptive might simply reflect passive responses to the environment, or represent by-products of adaptive plastic responses in other traits. In order to understand further the mechanism of plant shade tolerance, we therefore strongly recommend that future studies should explicitly test whether the plasticity of a trait is adaptive or not.

SUPPLEMENTARY DATA

Supplementary data are available online at www.aob.oxfordjournals.org and consist of the following. Methods S1: list of published studies from which data on plastic changes in biomass and SLA in response to shading were extracted. Methods S2: phylogenetic tree used in this study and list of published studies used for resolving the polytomies within the initial base tree. Table S1: mean effect size estimates from phylogenetically corrected meta-analytical models performed separately for each factor. Figure S1: relationship between plasticity in SLA and the changes in plant biomass when going from shade to high-light control environments, and plant biomass under different light conditions. Figure S2: means (\pm s.e.) of SLA in the high-light control treatments as estimated from a liner mixed-effects model for evergreen woody, deciduous woody and non-woody (herbaceous) species. Figure S3: boxplots of SLA values and biomass response to shading of species of different shade

intolerance classes. Figure S4: the relationship between plant biomass reduction and relative light intensity for 70 species that were grown under more than two light conditions. Figure S5: means of parameter estimates describing the relationship between biomass responses to shading and SLA plasticity in response to shading, SLA in the high-light control treatment, relative light intensity and type of experiment on the changes of plant biomass in response to shading.

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