Intestinal steroidogenesis

Guillaume Bouguen\textsuperscript{a,b,c}, Laurent Dubuquoy\textsuperscript{d,e}, Pierre Desreumaux\textsuperscript{d,e,f}, Thomas Brunner\textsuperscript{g}, Benjamin Bertin\textsuperscript{d,e,h,*}

\textsuperscript{a} Service des Maladies de l’Appareil digestif, CHU Pontchaillou, Rennes, France
\textsuperscript{b} UMR991, Liver Metabolism and Cancer, France
\textsuperscript{c} Université de Rennes 1, France
\textsuperscript{d} Université de Lille, F-59000 Lille, France
\textsuperscript{e} Inserm U995, F-59045 Lille, France
\textsuperscript{f} CHU Lille, Service des Maladies de l’Appareil Digestif et de la Nutrition, Hôpital Claude Huriez, F-59037 Lille, France
\textsuperscript{g} Biochemical Pharmacology, Department of Biology, University of Konstanz, Germany
\textsuperscript{h} Faculté des Sciences Pharmaceutiques et Biologiques, F-59006 Lille, France

Correspondence author at: Inserm U995, Faculté de médecine, pôle recherche – 4ème étage, 1 place de Verdun, 59045 Lille cedex, France. Tel.: +33 3 20 97 42 08; fax: +33 3 20 97 42 32.

E-mail address: benjamin.bertin-2@univ-lille2.fr (B. Bertin).

Abstract

Steroids are fundamental hormones that control a wide variety of physiological processes such as metabolism, immune functions, and sexual characteristics. Historically, steroid synthesis was considered a function restricted to the adrenals and the gonads. In the past 20 years, a significant number of studies have demonstrated that steroids could also be synthesized or metabolized by other organs. According to these studies, the intestine appears to be a major source of \textit{de novo} produced glucocorticoids as well as a tissue capable of producing and metabolizing sex steroids. This finding is based on the detection of steroidogenic enzyme expression as well as the presence of bioactive steroids in both the rodent and human gut. Within the intestinal mucosa, the intestinal epithelial cell layer is one of the main cellular sources of steroids. Glucocorticoid synthesis regulation in the intestinal epithelial cells is unique in that it does not involve the classical positive regulator steroidogenic factor 1 (SF 1) but a closely related homolog, namely the liver receptor homolog 1 (LRH 1). This local production of immunoregulatory glucocorticoids contributes to intestinal homeostasis and has been linked to pathophysiology of inflammatory bowel diseases. Intestinal epithelial cells also possess the ability to metabolize sex steroids, notably estrogen; this mechanism may impact colorectal cancer development. In this review, we contextualize and discuss what is known about intestinal steroidogenesis and regulation as well as the key role these functions play both in physiological and pathological conditions.

1. Introduction

In vertebrates, the mucosal surfaces provide a barrier against invading pathogenic microorganisms but represent also an essential interface exchange between the host and its environment \cite{1}. With a surface of broadly 300 m\textsuperscript{2} in adults, the intestinal mucosa is the biggest mucosal surface in humans. The gastrointestinal tract is constantly colonized by a huge number of microorganisms (approximately 100 trillion), mostly bacteria, but also viruses and fungi, which are collectively referred to as the microbiota \cite{2}. This colonization leads to a complex mutualistic relationship between the host and the microbiota in which each partner contributes positively to various physiological processes of the other one. For example, the microbiota digests and ferments carbohydrates that provide nutrients to the host cell, produces essential vitamins, contributes to the development of the gut immune system, and protects the host from potential pathogens. In turn, the host regulates microbial ecology by providing nutrients and ecological niches. It also controls microbial density through the synthesis of antimicrobial peptides and immunoglobulins. This dialog is required to maintain what is commonly called intestinal homeostasis. The disruption of this homeostasis may lead to the development of pathologies such as inflammatory bowel diseases \cite{3,4}.

A fundamental actor in the maintenance of the gut immune homeostasis are the intestinal epithelial cells (IECs) \cite{5,6}. IECs, located in the intestinal mucosa, provide a physical and biochemical barrier that separates microbiota and immune cells.
addition, interactions of IECs with the microbiota or with pathogens result in anti-microbial and immunoregulatory responses through the production of numerous molecules [5,7,9]. One immunoregulatory molecule produced locally by IECs is represented by glucocorticoids (GCs; i.e. cortisol in humans and corticosterone in rodents).

Historically, corticosteroid (glucocorticoids and mineralocorticoids) synthesis only occurred in the adrenal cortex. A significant number of studies have now challenged this view by demonstrating that organs such as the thymus [10-12], the skin [13], the brain [14-16], the intestine [17,19], and the lung [20] are capable of producing glucocorticoids.

This review aims to present evidence of local steroid synthesis, particularly that of GCs in the intestine, as well as the key factors involved in the regulation of this production. We discuss the known and potential roles of GCs in the regulation of intestinal homeostasis and then review the data showing that IECs, aside from their ability to produce GCs, are also involved in the metabolism of other steroids (notably sex steroids).

2. Intestinal glucocorticoids

2.1. Evidence of intestinal GC production

The steroidogenic ability of the gut was first suggested in the beginning of the 90’s after looking for the expression and activity of the CYP17 (P450c17) enzyme along the gastrointestinal tract of adult rats (see Fig. 1 for a schematic representation of the cortisone and sex steroids synthesis pathway). When Dalla Valle et al. assessed the ex vivo metabolism of [4-14C]pregnenolone in various organs, they found that the duodenum and the colon were able to produce high amounts of dehydroepiandrosterone (DHEA), attesting the presence of 17x-hydroxylase and C 17,20 lyase activities in the intestine [21,22]. While rodents are corticosterone secreting animals, due to the lack of expression of CYP17 in the adrenal, CYP17 expression in peripheral tissues (notably in the intestine) could lead to the synthesis of local cortisol in rats or mice. However, an early study failed to identify GCs (corticosterone or cortisol) in the small intestine of rats by thin layer chromatography [23]. Moreover, the hypothesis of locally produced cortisol in rodents has never been tested and only corticosterone has been investigated.

In the same period of time, the identification of the transcripts encoding both CYP11A1 (cholesterol side chain cleavage cytochrome P450; P450scc) and 3βHSD (3 beta hydroxysteroid dehydrogenase) in the primitive gut of the mouse embryo suggested the ability of the gut to synthesize steroids de novo [24]. The expression of 11βHSD2 (11 beta hydroxysteroid dehydrogenase 2), which converts active GCs to inactive metabolites, was also detected in the intestine of rodents and humans [25,26]. Subsequent studies demonstrated steroidogenic enzyme expression as well as intestinal steroid metabolism in various species such as frog [27], fish [28], mouse [17], and human [18,19,29]. Most of the comprehensive studies on intestinal steroidogenesis have been undertaken using mouse models. Cima et al. first measured (by radioimmunoassay) the de novo synthesis of corticosterone in murine small intestinal tissue fragments cultured ex vivo [17]. This production was highly stimulated in response to immune factors.
activation of T lymphocytes. Corticosterone synthesis was proven to be in situ, since GC release from tissue was blocked by metyrapone, a1-hydroxylase (P450c11p; CYP11B1) inhibitor, and 11β-hydroxylase (11βHSD) enzymes were all expressed in the murine small and large intestinal mucosa, but some of them were only detected after immune stimulation [17]. The cellular source of steroidogenic enzymes and GCs was shown to be the IECs located in the crypt region of the small intestine [17]. Further study demonstrated that the expression patterns of mRNA encoding CYP11A1 and CYP11B1 were linked to cell cycle. This restricts the main intestinal GC source to the proliferative cells of the intestinal crypts [32].

Initial work by Cima et al. suggested that GC synthesis could also occur in the human intestine, although results were not shown [17]. Earlier observations suggested intestinal steroid metabolism in humans following the detection of 11βHSD and 17 β hydroxy steroid dehydrogenase (17βHSD) enzymes in the colon and along the gastrointestinal tract [26,33]. More recently, CYP11A1, CYP17 and CYP11B1 and a production of cortisol were observed in colorectal cancer cell lines (Caco 2, HT 29) as well as in primary human intestinal tissue and colorectal tumors [18]. The underlying reasons of the discrepancy between adrenal and intestinal cells during the regulation of cortisol synthesis remain currently unknown and the control of intestinal steroidogenesis by one or several hormonal signals needs to be characterized.

The differential regulation of glucocorticoid synthesis in adrenal and intestinal cells is also illustrated by the observation that ACTH administration was able to trigger the release of corticosterone from the adrenal but not from the intestine [40]. ACTH mainly mediates its effect through the stimulation of adenylate cyclase leading to an increase of intracellular cyclic adenosine monophosphate (cAMP) level and subsequent protein kinase A activation. Thus, it was observed that increasing intracellular cAMP level in the adrenocortical tumor cell line Y1 induces the expression of CYP11A1 and CYP11B1, while the opposite effect was obtained in the murine intestinal epithelial cell line mIC12 [40]. Conversely, it was shown that phorbol ester dependent protein kinase C activation was involved in the control of the steroidogenic enzyme expression and GC production in mIC12 cells but not in Y1 cells [40]. However, LRH 1 mRNA level was slightly modified by cAMP and phorbol ester in mIC12. Recently Lee and colleagues observed in Hela cells the ability of phorbol ester to stimulate the phosphorylation of LRH 1 in an ERK1/2 dependent manner [44]. Therefore, rather than a modification of LRH 1 expression, different phosphorylation states of LRH 1 may explain the antagonistic effect of cAMP and phorbol ester observed in intestinal epithelial cells [40]. The underlying reasons of the discrepancy between adrenal and intestinal cells during the regulation of cortisol synthesis remain currently unknown and the control of intestinal steroidogenesis by one or several hormonal signals needs to be characterized.

To date, only the induction of strong immune responses has been found to induce intestinal steroidogenesis. Pioneering works have demonstrated that de novo synthesis of corticosterone from small intestinal tissue fragments was highly stimulated in response to immune activation of T lymphocytes [17]. Using lipo polysaccharide (LPS) injection in mice, subsequent studies investigated whether activation of the innate immune system could also induce intestinal steroidogenesis. LPS was able to induce intestinal CYP11A1 and CYP11B1 expression level and intestinal corticosterone release in Rag2-/- mice (which lack T and B lymphocytes), suggesting that intestinal GC synthesis induction also occurs independently of the activation of the adaptive immune system [43]. The authors showed that the cells, which contribute to this LPS induced intestinal steroidogenesis, were macrophages [43].

Finally, the pro inflammatory cytokine tumor necrosis factor (TNF) was clearly demonstrated to be the common mediator that regulates intestinal GC production in response to the activation of both the innate and adaptive immune system [43]. Following immune stimulation by LPS or activation of T cells, intestinal GC synthesis was compromised both in mice lacking TNF cytokine or TNF receptor expression. The involvement of TNF was later confirmed but the precise role of this cytokine needs further clarifications since opposite effects on gut steroidogenesis were found [45,46]. One might argue that the differential involvement of LRH 1 according to the immune response may explain such discrepancies. In wild type mice, both activated T cells and LPS were able to induce CYP11B1 expression and corticosterone production in a TNF dependent manner, but only the T cell dependent induction of intestinal steroidogenesis was compromised in LRH 1 haplodeficient animals (LRH 1+/−), which suggest that LPS drives intestinal GC production independently of LRH 1 [43]. By contrast,
Huang et al. suggested that TNF dependent pro inflammatory sig-
nals mediated by NF kB and JNK suppressed CYP11A1 and
CYP11B1 expression by inhibiting LRH 1 transcriptional activity
under chronic inflammatory conditions [46]. In summary, the
understanding of the regulatory mechanisms of GC synthesis in
the gut is just at the beginning, and additional mechanisms
involved in the regulation of intestinal steroidogenesis remain to
be discovered.

2.3. Function and roles of intestinal GCs

2.3.1. Maturation and maintenance of the intestinal epithelial barrier

GC mediated effects on the maturation of the small intestine are
well recognized both in rodents and humans [47]. For instance,
adrenalectomy in rats during the weaning period impairs the mor-
phological development of the small intestine [48], while injection
of cortisone or hydrocortisone to suckling rats increases the expres-
sion and activity of many small intestinal maturation markers
[49 51]. The development of a human intestinal xenograft model
demonstrated the responsiveness of the human small intestine to
cortisone during the maturation period [52,53]. More recently,
in vitro studies demonstrated the role of GCs in the maintenance
of intestinal epithelial barrier integrity [54]. Boivin et al. used a
human intestinal epithelial monolayer model consisting of filter
grown Caco 2 cells to investigate the role of GCs on intestinal bar-
rier function. By measuring transepithelial electric resistance
(TEER) and paracellular permeability, they showed that synthetic
GCs (prednisolone and dexamethasone) had a protective effect
against the TNF dependent increase of intestinal permeability
[54]. In vitro, a positive effect of dexamethasone on epithelial bar-
rier function was subsequently found independent of the presence
of any immunological stimulus [53]. The overall in vitro response of
human intestinal epithelial cells to GCs was also studied using
microarray analysis. This work showed that hydrocortisone differ-
entially regulates the expression levels of protein encoding genes
and enterocyte markers involved in polarization and tight junction
formation [56]. Altogether, these studies underlined that both
endogenous (adrenal derived) and exogenous GCs play a role dur-
ing the functional maturation of the gut and the maintenance of
intestinal epithelial barrier function. However, during these pro-
cesses the effective contributions of GCs produced locally by the
intestine remain to be established. As compared to the adrenal,
the amount of GCs synthesized by IECs under steady state condi-
tions is rather low, and to date there is no experimental evidence
of the impact of intestinal GCs on intestinal barrier maturation
and/or maintenance. It was previously reported that the specific
deletion of LRH 1 in mouse IECs disrupted intestinal steroidogene-
sis and rendered the conditional knockout animals more suscepti-
bile to experimental colitis [29], see Section 4.1. Although a
change of intestinal permeability due to the LRH 1 dependent
decrease of intestinal GC could explain this phenotype, this hypoth-
esis remains to be tested.

Using a short hairpin RNA strategy, we recently created a stable
Caco 2 cell line in which the expression of LRH 1 was reduced by
80% compared to a control cell line [19]. This downregulation of
LRH 1 resulted in a reduction of cortisol synthesis and release.
However, preliminary experiments did not observed any change of
the TEER of this stable cell line up to 21 days when it was differ-
entiated and grown as a monolayer on filters, suggesting that
intestinal derived cortisol may not participate in the development
of intestinal barrier (author’s unpublished data). Further experi-
ments are needed to investigate the potential role of intestinal
GCs on gut epithelial differentiation and barrier function, e.g. the
measurement of intestinal permeability of LRH 1 deficient mice
under steady state and inflammatory conditions.

2.3.2. Impact on innate and adaptive immune responses

Consistent with the general immunoregulatory properties of GC
[57,58], as well as the induction of intestinal steroidogenesis fol-
wowing immune activation, it is likely that local GC release is
involved in the regulation of intestinal immune responses. In order
to specifically appreciate the role of GCs released by extra adrenal
sources (such as the intestine), Cima and colleagues removed adre-
nal glands, and challenged these mice with various T cell depen-
dent immunological stresses [17]. Adrenalectomized animals
were notably infected with lymphocytic choriomeningitis virus
(LCMV) (which triggers a robust cytotoxic T lymphocyte response)
in the presence or absence of the GC synthesis inhibitor metyra-
pone. Then, the virus specific T cell activation in the intestine
was assessed by monitoring membrane markers expression and
cytokine production. LCMV infected and metyrapone treated adre-
alectomized mice displayed a higher T cell activation compared to
LCMV infected animals only, indicating that de novo synthesized
extra adrenal corticosterone inhibited the T cell response. Al-
though this work did not formally demonstrate the intestine as
the only source of GCs, results obtained by Cima et al. strongly sug-
gested its impact. [42]. Regarding specifically the gut, heterozygous
LRH 1+/− mice, which present a decreased expression of steroido-
genic enzymes and corticosterone release were more susceptible
to experimental colitis induced by either 2,4,6 trinitrobenzene
sulfonic acid (TNBS) or dextran sodium sulfate (DSS) [29]. How-
ever LRH 1 expression was reduced in all cell types of rodents, making
it difficult to exclude the contribution of GCs produced by other
cells, such as immune cells [59]. But the immunoregulatory role
of GCs synthesized by the IECs was further supported by more spe-
cific experiments in the same study when enterocyte specific LRH
1 deficient mice produced less colonic corticosterone after TNBS
1 treatment and displayed more severe inflammatory lesions in
the large intestine [29]. Beside the immunoregulatory properties
of GCs, other mechanisms such as the maintenance of the intestinal
barrier and/or the recovery of the epithelial layer by promoting
progenitor cell proliferation through LRH 1 might also regulate
colitis. However, chemically induced models of colitis imperfectly
reproduced the cellular and molecular aspects of the physiopath-
ology of inflammatory bowel diseases (IBD), these results led also
to the assessment of the expression of LRH 1 and intestinal GCs syn-
thesis in patients suffering from IBD (see Section 4.1).

Apart from the characterization of T lymphocytes as targets of
intestinal GCs, no other cellular target has been described yet. It
has recently been demonstrated that a fundamental dialog
between IEC and dendritic cells (DCs) takes place within the intes-
tinal mucosa to maintain gut homeostasis. DCs are professional
antigen presenting cells (APC) viewed as one of the gate keepers
of the immune system forming a bridge between the innate and
adaptive immune responses. Recent studies demonstrated that
IECs educate intestinal DCs to suppress inflammation and promote
tolerance and gut immune homeostasis [7,9,60]. Several immuno-
regulatory molecules, such as TGF β (transforming growth factor
β), retinoic acid or TSLP (thymic stromal lymphopoietin), produced
by IECs are crucially involved in this tolerogenic process [5]. There-
fore, it is tempting to speculate that immunoregulatory GCs
released by IECs might have a similar role in promoting the devel-
oment of DCs (or others APC) with tolerogenic properties. All the
same, this hypothesis remains to be tested.

2.3.3. Other properties of intestinal GCs

Next to GR mediated effects regulation of GC sensitive genes,
there is also accumulating evidence for non genomic effects of
GCs. For example, GCs have been reported to mediate certain bio-
logical effects via interaction with cellular membranes, via cyto-
solic GR and via membrane bound GC receptors [61].
Some gut specific properties have been allocated to this intestinal steriodogenesis. Cortisol released by IECs has been recently demonstrated to be involved in the control of the intestinal expression of PPAR\(\gamma\) (peroxisome proliferator activated receptor gamma), a key factor in the regulation of gut homeostasis. The colon is a major PPAR\(\gamma\) expressing tissue with the colonic epithelial cells as the main source of expression [62,63]. PPAR\(\gamma\) belongs to the nuclear receptor superfamily and has been shown to be a critical regulator of inflammatory responses [63,67]. This property involves typical positive gene regulation by binding of the PPAR\(\gamma\) RXR (RXR, retinoid X receptor) heterodimer to the PPAR response element (PPRE) located in the promoter of target genes. In addition a direct interference with transcription factors involved in the inflammatory response, such as NF \(\kappa\)B (nuclear factor kappa light chain enhancer of activated B cells), AP 1 (activating protein 1) and ROR\(\gamma\)t (retinoic acid receptor related orphan receptor gamma t), has been demonstrated in a mechanism often referred to as “transrepression” [67]. In a stable Caco2 cell line with effective LRH 1 knockdown cortisol production and release, as well as PPAR\(\gamma\) mRNA and protein expression were drastically reduced [19]. This inhibition of PPAR\(\gamma\) expression was reversed to the normal levels following the treatment of the cell line by exogenous GCs. As a control, knockdown of PPAR\(\gamma\) by the same strategy did not result in a decrease of cortisol release or LRH 1 expression [19]. These experiments pointed out a key role of intestinal cortisol synthesis in the regulation of intestinal PPAR\(\gamma\) and PPAR\(\gamma\) mediated gut homeostasis.

Environmental stresses induce adaptive response of IECs with high secretory activity. The endoplasmatic reticulum (ER) folds and matures most secreted and transmembrane proteins. The imbalance between protein load observed in stress situation and folding ability of the ER is defined as ER stress, and the coordinated response triggered is known as UPR (Unfold Protein Response) to ensure homeostatic cell control [68,69]. UPR induces several pathways that lead to a reduced protein load that enters the ER, to increase the ability of the ER to handle unfolded proteins and in some circumstances to trigger cell death [68]. In the intestine dis regulation of normal ER responses are particularly sensitive since dysfunction of UPR leads to spontaneous colitis in rodents lacking the transcription factor XBP1 (X box binding protein 1), involved in protein load [70,71]. In Winnie mice with increased intestinal ER stress caused by a missense mutation of the Muc2 mucin gene, GCs suppress ER stress and activation of the UPR in a glucocorticoid receptor dependent manner [72]. GCs promote correct folding of secreted proteins and facilitate the enhanced removal of misfolded proteins from the ER through up regulation of genes encoding chaperones and elements of ER associated degradation (ERAD) [72]. These results suggest that epithelial steriodogenesis may also influence gut homeostasis by modulating ER responses in response to exogenous and endogenous stresses.

3. Metabolism and function of other steroids in the gut

The steriodogenesis involves not only the production of GCs but also other steroids such as mineralocorticoids and sex steroids (androgens and estrogens). In parallel to the description of gut production of GCs, the ability of IECs to metabolize sex steroids was assessed. From a chemical point of view, few enzymatic steps are required to obtain androgens and estrogens from common precursors of GCs involving the 3 enzymes 17\(\alpha\) hydroxylase/17,20 lyase (P450c17), and the 17\(\beta\) hydroxysteroid dehydrogenase (17\(\beta\)HSD).

For estrogens there is an additional enzyme, the aromatase (P450aro) (Fig. 1). Early in vitro and in vivo studies observed the expression of enzymes involved in estrogen metabolism in colorectal cancer cell lines and normal gut mucosa [33,73,76]. Evidence of estrogen production by the IECs in healthy humans was then assessed leading to the observation of higher mucosal production of estrone as compared to estradiol [77].

An important aspect of estrogens is their synthesis in diverse tissues. This may lead to variable exposition of IECs by estrogens, depending on sex or diet, in addition to their own production in the intestinal mucosa. The 17\(\beta\) hydroxysteroid dehydrogenase (17\(\beta\)HSD) family acts in key steps of the synthesis of sex steroid hormones, mainly of active estrogens and androgens [78]. Several isoforms of the 17\(\beta\) HSD enzyme with diverse enzymatic activities have been characterized in humans and were thoroughly investigated in human colonic mucosa, which mainly express isoform 2 and 4 [75]. These isoforms catalyze the oxidative transformation of estradiol and testosterone to estrone and androstenedione, respectively, which is thus considered to play an important role in the peripheral inactivation of estrogen and androgen [33,74,78,79].

Similarly to GCs, estrogens metabolism (mainly estradiol) seems to occur predominantly in the intestinal crypts of the colon and may thereby exert a significant effect on colonic epithelial cell growth [33,80]. Of interest in this regard is the observation that ovarectomy appears to induce colonic crypt atrophy [81]. Estrogen receptor \(\beta\) (ER \(\beta\)) is highly expressed in gut epithelial cells [82,83] and ER \(\beta\) signaling is involved in cellular homeostasis and maintenance of the colonic epithelial barrier function [84,85]. Disruption of ER \(\beta\) expression in ER \(\beta\) KO mice was notably associated with an increase in epithelial cell proliferation, and a decrease of apoptosis and the expression of cellular adhesion molecules [84].

4. Involvement of intestinal steriodogenesis in pathological disorders

4.1. Inflammatory bowel diseases

Inflammatory bowel diseases (IBDs), namely ulcerative colitis (UC) and Crohn’s disease (CD) are chronic, complex, relapsing immune mediated disorders of the gastrointestinal tract of unknown etiology. Emerging evidence suggests that disease development involves a deregulated immune response to the gut microbiome in genetically susceptible individuals, triggered by environmental factors [86,89]. The expression of CYP11A1, CYP11B1 and LRH 1 were first shown to be decreased in inflamed genetic susceptible colonic mucosa in in vitro and in vivo experiments including in surgical specimens of UC patients and control subjects [29]. These observations strongly suggest that impaired production of intestinal GCs might participate in the pathogenesis of IBD. This notion was recently newly strengthened by our recent report [19]. While UC and CD share some clinical manifestations, they are distinct pathophysiological diseases. For instance, in contrast to the characteristic transmural lesions that can affect all the intestinal layers in CD patients, inflammation during UC is restricted to colonic mucosa and sub mucosa. Moreover, recent studies suggest that IEC dysfunction may critically contribute to the pathogenesis of UC [90,91]. Our recent work demonstrated that colonic epithelial cells (CEC) isolated from surgical specimens of UC patients compared to non inflamed biopsies of IBD patients and control subjects [29]. These observations strongly suggest that impaired production of intestinal GCs might participate in the pathogenesis of IBD. This notion was recently newly strengthened by our recent report [19]. While UC and CD share some clinical manifestations, they are distinct pathophysiological diseases. For instance, in contrast to the characteristic transmural lesions that can affect all the intestinal layers in CD patients, inflammation during UC is restricted to colonic mucosa and sub mucosa. 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production of GCs is a key element in the control of intestinal homeostasis and that the defective synthesis of cortisol by IECs may actively contribute to the pathogenesis of UC.

During IBD two other circumstances may involve intestinal steroidogenesis. We discussed above the possible impact of GCs on ER stress. Induction of ER stress in intestinal epithelium through tissue (and cell type) specific disruption of XBPI was shown to result in spontaneous enteritis due to inability of XBPI deficient IECs to properly generate antimicrobial activity and respond appropriately to inflammatory signals in the local milieu [70]. This concept was translated to patients with IBD, who are more likely to develop both CD and UC when bearing single nucleotide polymorphisms within the XBPI gene locus [70]. The steroidogenesis state in patients with CD is not yet known. Since GCs are able to relieve the ER stress [72], UPR are more likely to be impaired in UC patients with defective IEC synthesis of cortisol.

Another side of steroidogenesis implies sex hormones, particularly estrogens. In two large prospective cohorts of women a significant association between oral contraceptive use and mainly 17HSD isoform 2 [77,79]. The 17HSD2 catalyzes the oxidative transformation of estradiol to estrone [74]. Both the level of estradiol and 17\text{HSD} have been associated with CRC and distant progression irrespective of sex. Since IECs are a source of estrogen, the control of epithelial steroidogenesis could be useful to control CRC onset and progression.

Another hypothetical involvement of IEC steroidogenesis may be via PPAR\gamma. Beside the anti-inflammatory properties of PPAR\gamma, this nuclear receptor regulates cell proliferation, differentiation and is able to induce apoptosis [95]. Activation of PPAR\gamma by a synthetic ligand (mescaline) and contraceptive hormones is associated with a decrease in CD, were also associated with a modest, but non significant increase in risk of UC [92]. Conversely postmenopausal hormone use was associated with an increased risk of UC but not CD. As discussed previously, estrogen signaling is involved in the colonic barrier function [84,85]. Decreased estrogen receptor \beta mRNA levels were observed in colonic biopsies from IBD patients as well as in IL 10 deficient mice (a model of IBD), which correlated with an increase in epithelial permeability [85]. These findings indicate a potential impact of sex hormones, in particular estrogens, on IBD development. However, to our knowledge no study has thus far assessed estrogen synthesis in the intestinal mucosa of IBD patients.

4.2. Colorectal cancer (CRC)

Epidemiological and observational studies pointed out a correlation of exogenous and endogenous estrogens in the development of colorectal cancer (CRC) with a possible increased risk in women with higher circulating endogenous estrogen concentrations [93]. Estrogens may induce colorectal cancer cell apoptosis by binding to estrogen receptor \beta, [73,94]. A2.4 fold increased of estrogen (mainly estrone rather than estradiol) were observed in colon carcinoma tissues [77]. This may be related to overexpression of enzymes involved in steroid synthesis pathway, such as sulfatase and mainly 17HSD isoform 2 [77,79]. The 17\text{HSD2} catalyzes the oxidative transformation of estradiol to estrone [74]. Both the level of estradiol and 17\text{HSD} have been associated with CRC and disease progression irrespective of sex. Since IECs are a source of estrogen, the control of epithelial steroidogenesis could be useful to control CRC onset and progression.

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5. Conclusion

Steroid metabolism has been demonstrated and confirmed in the intestine. IECs are able to de novo synthesize and release GCs that regulate the gut immune response. The defective synthesis of cortisol identified in the colonic epithelial cells of UC patients strengthens the idea that intestinal GCs contribute to intestinal homeostasis. In addition, perturbations of estrogen metabolism are also evident in CRC cells, strongly suggesting that steroid metabolism is an important process during IEC homeostasis.

Whereas in classical steroidogenic organs (adrenals and gonads), the regulation of steroid production is well characterized, a lot of work remains to be done in order to identify the mechanisms controlling intestinal steroidogenesis. For instance, the recent identification of microbiota as a potential regulator of corticosterone production by the small intestine in mice [98] may pave the way to design new studies that aim to more thoroughly characterize the production of GCs in the gut. Given the involvement of steroids produced by the gut in pathological conditions such as CRC and IBD, therapeutic strategies targeting intestinal steroidogenesis might represent interesting new approaches for the treatment of these chronic diseases.

Conflict of interest

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71